

# TMV 296 - IRON SULPHITE AGAR (VEG.)

## **INTENDED USE**

For detection of thermophillic anaerobic organisms causing sulphide spoilage in foods.

### **PRODUCT SUMMARY AND EXPLANATION**

This medium is prepared by using veg hydrolysate which is free from BSE/TSE risks associated with animal based peptones. Iron Sulphite Veg Agar is the modification of Iron Sulphite Agar which is based on formula of Cameron Sulphite Agar developed by the National Canners Association of America. 0.1% sulphite concentration in the original formula was reported by Beerens to be inhibitory to some strains of *Clostridium sporogenes*. This observation was later on confirmed by Mossel et al, who consequently showed that 0.05% sulphite concentration was not inhibitory to the organisms. For the detection of organisms causing sulphide spoilage either Deep-Shake Culture method or Attenborough and Scarr method may be followed:

In Deep-Shake Culture method, the medium is dispensed in 10 ml amounts in sterile tubes. Sample is inoculated when the medium is at about 50°C and allowed to set. Further incubation is carried out at 55°C for 24-48 hours. Typical thermophilic species of *Desulfotomaculum nigrificans*, produces distinct black spherical colonies in the depth of the medium.

In Attenborough and Scar method, diluted samples of sugar or any other food to be tested are filtered through membrane filters. These filters are then rolled up and placed in tubes containing just sufficient Iron Sulphite Veg Agar (at 50°C) to cover them. The medium is allowed to set and then incubated at 55 - 56°C for 24 - 48 hours. After incubation, the number of black colonies on the membrane filter are counted. This membrane filter technique is quicker, of comparable accuracy and permits the examination of larger samples.

# COMPOSITION

Ingredients	Gms / Ltr
Veg hydrolysate	10.0
Sodium sulphite	0. 5
Iron (III) Citrate	0. 5
Agar	15.0

#### PRINCIPLE

Veg hydrolysate provides protein, nitrogen and other nutrients necessary to support bacterial growth. Sulphite-reducing bacteria usually produce black colonies as a result of the reduction of sulphite to sulphide, which reacts with the iron (III) salt.

### **INSTRUCTION FOR USE**

- Dissolve 26 grams in 1000 ml distilled water.
- Heat to boiling to dissolve the medium completely.
- Sterilize by autoclaving at 15 psi pressure (121°C) for 15 minutes.

QUALITY CONTROL SPECIFICATIO	NS
Appearance of Powder	: Yellow coloured, may have slightly greenish tinge, homogeneous, free flowing powder.
Appearance of prepared medium pH (at 25°C)	: Yellow coloured, slightly opalescent gel forms in petri plates. : 7.1±0.2

## **INTERPRETATION**

A- 902A, RIICO Industrial Area, Phase III, Bhiwadi-301019.



2

f 0 in 1



## Cultural characteristics observed under anaerobic conditions, after an incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Colour of colony	Incubation Temperature	Incubation Period
Clostridium botulinum	25763	50-100	Luxuriant	>=70%	Black	55-56°C	24-48 Hours
Clostridium sporogenes	19404	50-100	Luxuriant	>=70%	Black	55-56°C	24-48 Hours
Desulfotomacul um nigrificans	19998	50-100	Luxuriant	>=70%	Black	55-56°C	24-48 Hours
Escherichia coli	25922	50-100	Good	40-50%	No blackening	55-56°C	24-48 Hours

#### PACKAGING:

In pack size of 100 gm and 500 gm bottles.

## STORAGE

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

**Product Deterioration:** Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

# DISPOSAL

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

#### REFERENCES

- 1. Tanner F.W., 1944, "The Microbiology of Foods", 2nd ed., Garrard Press, Illinois, P. 1127.
- 2. Beerens H., 1958, DSIR, Proc. 2nd Internat. Sym. Food Microbiol., 1957, HMSO, London, P. 235.
- 3. Mossel D.A.A., Golstein Brouwers G.W.M.V. and de Bruin A.S., 1959, J. Path. Bact., 78:290.
- 4. Attenborough J. and Scarr M., 1957, J. Appl. Bact., 20:460.





NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices. \*For Lab Use Only Revision: 08 Nov., 2019

