

TMP 027GT -SOYABEAN CASEIN DIGEST AGAR PLATE W/ 1% GLYCEROL,0.5% LECITHIN & 4% POLYSORBATE 80 (g - irradiated) (Triple Pack)

INTENDED USE

For determining of sanitization of containers, equipment, surfaces, water miscible cosmetics etc.

PRODUCT SUMMARY AND EXPLANATION

Soyabean casein Digest Agar Plate w/ 1% Glycerol, 0.5% Lecithin and 4% Polysorbate 80 plates are recommended for the isolation of microorganisms from environmental surfaces and is used primarily to monitor microbial contamination, enumerate the number of microbial colonies growing on a variety of surfaces sanitized with quaternary ammonium compounds, and to assist in determining surface sanitation.

COMPOSITION

Ingredients	Gms / Ltr
Polysorbate 80 (Tween 80)	40.000
Agar	15.000
Casein enzymic hydrolysate	15.000
Glycerol	10.000
Papaic digest of soyabean meal	5.000
Sodium chloride	5.000
Lecithin	5.000

PRINCIPLE

Medium contains Casein enzymic hydrolysate and papaic digest of soyabean meal which helps to provide nitrogenous compounds and other nutrients essential for microbial replication. Sodium chloride is added to maintain cellular osmotic equilibrium. Lecithin and polysorbate 80 are added to the formulation to neutralize germicidal or disinfectant residues. Neutralization of these residues reduces their inhibitory effect which ultimately results in lowering of microbial count. Quaternary ammonia compounds are neutralized by lecithin, while phenolic disinfectants and hexachlorophene are neutralized by polysorbate 80. Together, lecithin and polysorbate 80 neutralize ethanol.

INSTRUCTION FOR USE

Either streak, inoculate or surface spread the test inoculum aseptically on the plate. Alternatively, these plates can also be used as contact plates for environmental monitoring.

QUALITY CONTROL SPECIFICATIONS

Appearance	: Light to medium amber colour medium.
Quantity of Medium	: 30±2 ml of medium in 90 mm sterile disposable Petri-plates.
pH (at 25°C)	: 7.3± 0.2
Dose of irradiation:	: 15-25 kGy
Sterility Check	: Passes release criteria

INTERPRETATION



Cultural characteristics observed after incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Incubation Temperature	Incubation Period
<i>Escherichia coli</i>	25922	50-100	Luxuriant	≥70 %	35-37 °C	18-24 Hours
<i>Staphylococcus aureus</i>	25923	50-100	Luxuriant	≥70 %	35-37 °C	18-24 Hours
<i>Pseudomonas aeruginosa</i>	27853	50-100	Luxuriant	≥70 %	35-37 °C	18-24 Hours
<i>Bacillus subtilis</i>	6633	50-100	Luxuriant	≥70 %	35-37 °C	18-24 Hours
<i>Salmonella typhimurium</i>	14028	50-100	Luxuriant	≥70 %	35-37 °C	18-24 Hours
<i>Candida albicans</i>	10231	50-100	Luxuriant	≥70 %	30-35 °C	48-72 Hours

PACKAGING:

Triple layered packing containing 5 No. of plates with one silica gel desiccant bag packed inside it.

STORAGE

On receipt, store the plates at 15–30 °C. Avoid freezing and overheating. Do not open until ready to use. Prepared plates stored in their original sleeve wrapping until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation times. Allow the medium to warm to room temperature before inoculation.

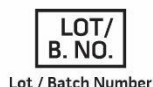
Product Deterioration: Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

DISPOSAL

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES

1. The United States Pharmacopoeia. 2009. Amended Chapters 61, 62 & 111, The United States Pharmacopoeial Convention Inc., Rockville, MD.
2. Directorate for the Quality of Medicines of the Council of Europe (EDQM). 2009. The European Pharmacopoeia, Amended Chapters 2.6.12, 2.6.13, 5.1.4, Council of Europe, 67075 Strasbourg Cedex, France.
3. Japanese Pharmacopoeia. 2008. Society of Japanese Pharmacopoeia. Amended Chapters 35.1, 35.2.7. The Minister of Health, Labor, and Welfare.
4. Indian Pharmacopoeia. 2010. Govt. of India, Ministry of Health and Family Welfare, New Delhi, India.
5. MacFaddin, J.F. 1985. Media for isolation-cultivation-identification-maintenance of medical bacteria, vol. I. Williams & Wilkins, Baltimore.
6. Baron, E.J., L.R. Peterson, and S.M. Finegold. 1994. Bailey & Scott's diagnostic microbiology, 9th ed. Mosby-Year Book, Inc., St. Louis.
7. Chapin, K.C., and P.R. Murray. 1999. Media, p. 1687-1707. In P.R. Murray, E.J. Baron, M.A. Pfaller, F.C. Tenover, and R.H. Tenover (ed.), Manual of clinical microbiology, 7th ed. American Society for Microbiology, Washington, D.C.
8. Clesceri, L.S., A.E. Greenberg, and A.D. Eaton (ed.). 1998. Standard methods for the examination of water and wastewater, 20th ed. American Public Health Association, Washington, D.C.
9. Downes, F.P. and K. Ito. (ed.). 2001. Compendium of methods for the microbiological examination of foods, 4th ed. American Public Health Association, Washington, D.C.
10. ISO 11137-1: 2006 + Amd 1:2013. Sterilization of health care products – Radiation - Part 1: Requirements for the development, validation and routine control of a sterilization process for medical devices.
11. ISO 11137-2:2013. Sterilization of health care products -- Radiation -- Part 2: Establishing the sterilization dose.



NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.



*For Lab Use Only
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