TM 626 – PSEUDOMONAS AGAR BASE

INTENDED USE

For selective isolation of *Pseudomonas* species from environmental samples, food and water.

PRODUCT SUMMARY AND EXPLANATION

Pseudomonas Agar Base is a modification of Kings A medium which contains magnesium chloride and potassium sulphate to enhance pigment production. Goto and Enomoto formulated CetriNix supplement for the selective isolation of *Pseudomonas aeruginosa* from clinical specimens. Lowbury and Collins studied cetrimide as a selective agent. CetriNix supplement suppresses *Klebsiella*, *Proteus* and *Providencia* species.

C-F-C Supplement was formulated by Mead and Adams making the medium specific for isolation of *Pseudomonas* from chilled foods and processing plants, environmental samples and water. This medium is recommended for enumeration of *Pseudomonas* species from meat and meat products. It can also be used for clinical samples.

COMPOSITION

Ingredients	Gms / Ltr
Tryptone	10.000
Gelatin peptone	16.000
Potassium sulphate	10.000
Magnesium chloride, anhydrous	1.400
Agar	11.000

PRINCIPLE

This medium consists of Tryptone and gelatin peptone which supplies nitrogenous and carbonaceous compounds, long chain amino acids, and other essential growth nutrients.

INSTRUCTION FOR USE

- Dissolve 24.20 grams in 500 ml purified/distilled water containing 5 ml glycerol.
- Heat to boiling to dissolve the medium completely.
- Sterilize by autoclaving at 15 psi pressure (121°C) for 15 minutes.
- Cool to 45-50°C and aseptically add sterile rehydrated contents of either CetriNix Supplement or CFC Supplement as desired.
- Mix well and pour into sterile Petri plates.
  Note: Do not keep the molten agar for longer than 4 hours.

QUALITY CONTROL SPECIFICATIONS

Appearance of Powder : Cream to yellow homogeneous free flowing powder.

Appearance of prepared medium : Yellow coloured Clear to slightly opalescent gel forms in Petri plates.

pH (at 25°C) : 7.1 ± 0.2

INTERPRETATION

Cultural characteristics observed after incubation. Recovery rate is considered as 100% for growth on Soyabean Casein Digest Agar.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth (at 34-38°C with TS 075)	Recovery (at 34-38°C with TS 075)	Growth (at 24-26°C with TS 075)	Recovery (at 24-26°C with TS 077)	Colour/ Fluorescence under UV	Incubation Period
Proteus vulgaris	13315	>=10^3	Inhibited	0%	-	-	-	40-48 Hours
Pseudomonas aeruginosa	27853	50-100	Good-luxuriant	>=50%	-	-	Blue-green /positive	40-48 Hours
Pseudomonas aeruginosa	9027	50-100	Good-luxuriant	>=50%	-	-	Blue-green /positive	40-48 Hours
Pseudomonas aeruginosa	10145	50-100	Good-luxuriant	>=50%	-	-	Blue-green /positive	40-48 Hours
Pseudomonas cepacia	10661	50-100	-	-	Good-luxuriant	>=50%	-	40-48 Hours
Pseudomonas fluorescens	13525	50-100	-	-	Good-luxuriant	>=50%	-	40-48 Hours
Pseudomonas fragi	4973	50-100	-	-	Good-luxuriant	>=50%	-	40-48 Hours
Enterococcus faecalis	29212	>=10^3	Inhibited	0%	-	-	-	40-48 Hours
Enterococcus faecalis	19433	>=10^3	Inhibited	0%	-	-	-	40-48 Hours
Escherichia coli	25922	>=10^3	Inhibited	0%	Inhibited	0%	-	40-48 Hours
Escherichia coli	8739	>=10^3	Inhibited	0%	Inhibited	0%	-	40-48 Hours

PACKAGING:
In pack size of 100 gm and 500 gm bottles.

STORAGE
Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.
Product Deterioration: Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

DISPOSAL
After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES
1. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington, D.C.
2. Salfinger Y., and Tortorello M.L. Fifth (Ed.), 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
3. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1. HiMedia Laboratories
4. King E.O., Ward M.K. and Raney D.E., 1954, J.Lab and Clin. Med., 44:301.
5. Lowbury E.J. and Collins A.G., 1955, Clin. Path., 8:47.
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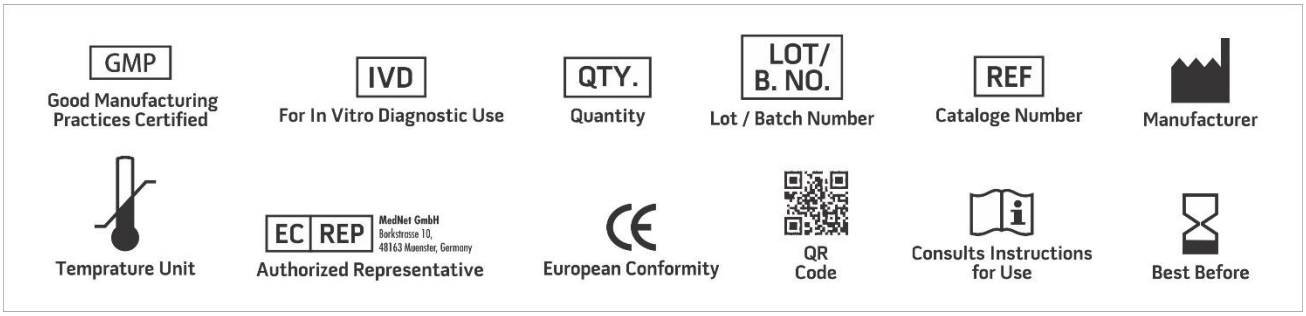

NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.
***For Lab Use Only**
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