

# TM 2417 - VIOLET RED BILE AGAR (1.2 %)

#### **INTENDED USE**

For selective isolation and enumeration of coli-aerogenes in water, milk and other dairy food products.

### PRODUCT SUMMARY AND EXPLANATION

Violet Red Bile Agar, a modification of MacConkeys original formulation is used for the enumeration of coli-aerogenes bacterial group. It relies on the use of the selective inhibitory components crystals violet and bile salts and the indicator system lactose, and neutral red. Thus, the growth of many unwanted organisms is suppressed, while tentative identification of sought bacteria can be made. Organisms, which rapidly attack lactose, produce purple colonies surrounded by purple halos. Non-fermenters or late lactose-fermenters produce pale colonies with greenish zones. VRBA is recommended by APHA. Violet Red Bile Agar (1.2 % Agar) is prepared, in accordance with the ISO Committee. Selectivity of VRBA can be increased by incubation under anaerobic conditions and/ or at elevated temperature, i.e. equal to or above 42°C.

### **COMPOSITION**

Ingredients	Gms / Ltr
Peptone	7.000
Yeast extract	3.000
Lactose	10.000
Bile salts mixture	1.500
Sodium chloride	5.000
Neutral red	0.030
Crystal violet	0.002
Agar	12.000

### **PRINCIPLE**

Peptone and yeast extract serve as sources of carbon, nitrogen, vitamins and other essential growth nutrients. Lactose is the fermentable carbohydrate, utilization of which leads to the production of acids. Neutral red indicator detects the acidity so formed. Crystal violet and bile salts mixture help to inhibit the accompanying gram-positive and unrelated flora. Sodium chloride maintains the osmotic equilibrium. Violet Red Bile Agar is not completely specific for enteric; other accompanying bacteria may give the same reaction. Further biochemical tests are necessary for positive identification.

## **INSTRUCTION FOR USE**

- Dissolve 38.53 grams in 1000 ml purified / distilled water.
- Heat with stirring to boiling to dissolve the medium completely, do not autoclave.
- Cool to 45-50°C and pour into sterile Petri plates containing the test sample.

# **QUALITY CONTROL SPECIFICATIONS**

**Appearance of Powder** : Light yellow to pink homogeneous free flowing powder.

**Appearance of prepared medium**: Reddish purple coloured clear to slightly opalescent gel forms in Petri plates.

**pH (at 25°C)** : 7.4±0.2

# **INTERPRETATION**

Cultural characteristics observed after an incubation.











Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Color of the colony	Incubation Temperature	Incubation Period
Klebsiella aerogenes	13048	50-100	Luxuriant	>=70%	Pink to pinkish	35-37°C	18-24 Hours
Escherichia coli	25922	50-100	Luxuriant	>=70%	Pinkish red With bile precipitate	35-37°C	18-24 Hours
Salmonella Enteritidis	13076	50-100	Luxuriant	>=70%	Colourless to Orangish yellow	35-37°C	18-24 Hours
Staphylococcus aureus subsp.aureus	25923	>=10 <sup>3</sup>	Inhibited	0%	-	35-37°C	18-24 Hours

#### **PACKAGING:**

In pack size of 500 gm bottles.

# **STORAGE**

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

Product Deterioration: Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

### **DISPOSAL**

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

#### REFERENCES

- 1. American Public Health Association, Standard Methods for the Examination of Dairy Products, 1978, 14th Ed., Washington D.C.
- 2. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington,
- 3. Corry J. E. L., Curtis G. D. W. and Baird R. M., (Ed.), 1995, Culture Media for Food Microbiology, Vol. 34, Progress in Industrial Microbiology, Elsevier, Amsterdam. International Organization for Standardization (ISO), 1991, Draft ISO/DIS 43
- 4. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Editio
- 5. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
- 6. MacConkey A., 1905, J. Hyg., 5, 333-3
- 7. Mossel D. A. A. and Vega C. L., 1973, Hlth. Lab. Sci., 11:3
- Mossel D. A. A., Eclderink I., Koopmans M. and Van Rossem F., 1979, Food Protect., 42: 4
- 9. Mossel D. A. A. et al, 1986, J. Appl. Bacteriol., 60:2
- 10. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
- 11. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.





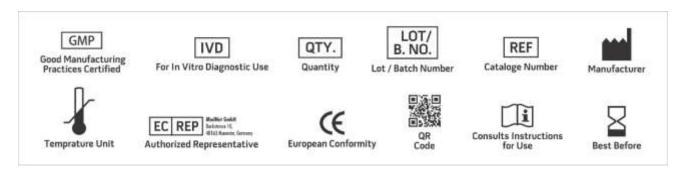












**NOTE:** Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices. \*For Lab Use Only Revision: 08 Nov., 2019







