

## TM 2383 – TRYPTONE LACTOSE IRON AGAR

### INTENDED USE

For identification of anaerobes on the basis of motility, hydrogen sulphide production and lactose fermentation.

### PRODUCT SUMMARY AND EXPLANATION

Tryptone Agar was developed by Vera for the accurate differentiation and identification of aerobes and anaerobes by means of motility and fermentation reactions. Tryptone Lactose Iron Agar medium is recommended to study motility of organism and simultaneous sulphite reduction in acidic environment. Due to presence of phenol red in the medium, on fermentation of lactose the medium turns yellow due to production of acid and gas. The ability of an organism to produce H<sub>2</sub>S is a consistent characteristics and an H<sub>2</sub>S producer usually produce gas (CO<sub>2</sub> + H<sub>2</sub>) in carbohydrate media which is visualized as air bubbles in the medium.

H<sub>2</sub>S production takes place in the presence of R1-SH group provided by cystine present in casein enzymic hydrolysate or through reduction of an inorganic sulphur source such as thiosulphate. H<sub>2</sub>S is a colourless gas, which upon contact with ferrous salt produces ferrous sulphide, a black precipitate indicated by a visible black reaction. Sodium sulphite at a concentration less than 0.05% is not inhibitory to *Clostridium sporogenes*.

### COMPOSITION

Ingredients	Gms / Ltr
Casein enzymic hydrolysate	20.000
Lactose	10.000
Ferrous sulphate	0.200
Sodium sulphite	0.400
Sodium thiosulphate	0.080
Phenol red	0.020
Agar	3.500

### PRINCIPLE

Casein enzymic hydrolysate provides essential growth nutrients to support the growth of organisms. Phenol red is the pH indicator. Even small amount of agar renders it suitable for study of motility. Small amounts of acid produced do not readily get dispersed throughout the medium and hence positive reaction can be more quickly determined in this medium than in liquid medium. Lactose is the fermentable carbohydrate.

### INSTRUCTION FOR USE

- Dissolve 34.2 grams in 1000 ml distilled water.
- Heat boiling to dissolve the medium completely. Dispense in test tubes.
- Sterilize by autoclaving at 12 psi 118°C for 15 minutes.
- Cool the tubes in an upright position.

### QUALITY CONTROL SPECIFICATIONS

Appearance of Powder	: Light yellow to light pink homogeneous free flowing powder.
Appearance of prepared medium	: Red coloured clear to slightly opalescent gel forms in tubes as butts.
pH (at 25°C)	: 7.3±0.2

### INTERPRETATION



Cultural characteristics observed after incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Acid	H <sub>2</sub> S	Motility	Incubation Temperature	Incubation Period
<i>Clostridium perfringens</i>	13124	50-100	Luxuriant	Positive reaction, yellow colour	Positive, blackening of medium	Positive, growth away from stabline causing turbidity	35-37°C	18-48 Hours
<i>Clostridium sporogenes</i>	11437	50-100	Luxuriant	Positive reaction, yellow colour	Negative, no blackening of medium	Positive, growth away from stabline causing turbidity	35-37°C	18-48 Hours

**PACKAGING:**

In pack size of 500 gm bottles.

**STORAGE**

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.













**Product Deterioration:** Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

**DISPOSAL**

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

**REFERENCES**

- 1.Vera, 1944, J. Bacteriol., 47:455.
- 2.MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
- 3.Clarke P. H. and Cowen S. T., 1952, J. Gen. Microbiol., 6:187.
- 4.Fieser L. F. ad Fieser M., 1956, Organic Chemistry, 3rd Ed., New York Reinhold Publishing Corporation. pg 155.
- 5.Doelle H. W., 1969, Bacterial Metabolism, New York, Academic Press, p. 99, 224.
- 6.Padron A. P. and Dockstader W. B., 1972, Appl. Microbiol., 23:1107. 7.Mossel D. A. A, et al, 1959, J. Path. Bacteriol., 78: 290.

 GMP Good Manufacturing Practices Certified	 IVD For In Vitro Diagnostic Use	 QTY. Quantity	 LOT/ B. NO. Lot / Batch Number	 REF Catalogue Number	 Manufacturer
 Temperature Unit	 EC REP Authorized Representative <small>MedNet GmbH Borkstrasse 10, 48163 Muenster, Germany</small>	 European Conformity	 QR Code	 Consults Instructions for Use	 Best Before

**NOTE:** Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.  
\*For Lab Use Only

Revision: 08 Nov., 2019

