

# TM 2090 - FOLIC ACID MEDIUM, AOAC

#### **INTENDED USE**

For the microbiological assay of folic acid using Enterococcus hirae ATCC 8043.

## PRODUCT SUMMARY AND EXPLANATION

Folic acid is required for the growth of *Enterococcus hirae*. Hence growth of this organism will occur only if the sample being assayed contains folic acid. The exact folic acid concentration in the test sample can be determined by comparing the growth obtained to that of known standard concentrations of folic acid (standard curve).

Folic Acid Assay Medium is prepared according to the formula described by Capps et al and is recommended by AOAC for the determination of folic acid content of the pharmaceutical products and other materials using *Enterococcus hirae* ATCC 8043 as the test organism.

Procedure: Stock cultures of *Enterococcus hirae* ATCC 8043 are prepared by stab inoculation of Lactobacilli Agar AOAC. Following incubation at 35-37°C for 24 hours, the tubes are stored in the refrigerator. Transplants are made at monthly intervals. Inoculum for assay is prepared by sub culturing from a stock culture of *Enterococcus hirae* ATCC 8043 into a tube containing 10 ml of Lactobacilli Broth AOAC. After 24 hours incubation at 35-37°C, the cells are centrifuged under aseptic conditions, and the supernatant liquid is decanted. The cells are resuspended in 10 ml of sterile 0.85% NaCl. The cell suspension is then diluted 1:100 with sterile 0.85% NaCl. One drop of this later suspension is used to inoculate each of the assay tubes. It is essential that a standard curve be set up for each separate assay since conditions of autoclaving, temperature of incubation, etc., which influence the standard curve readings cannot be duplicated exactly from time to time. The standard curve is obtained by using folic acid at levels of 0, 2, 4, 6, 8 and 10 ng per assay tube (10 ml). Tubes are refrigerated for 15-30 minutes to stop growth before reading. Turbidimetric readings should be read after 16-18 hours incubation at 35-37°C and Acidimetric after 72 hours at 35-37°C. To prepare stock solution of folic acid, 20 mg folic acid is used.

#### Preparation of Folic Acid Concentrations:

Dissolve 20 mg dried folic acid in 100 ml distilled water containing 20 ml ethanol. Adjust the pH of the solution to 10.0 with 0.1 N NaOH to dissolve the acid and then adjust pH to 7.0 with 0.05 N HCl. This solution contains 200 mcg folic acid per ml. Dilute 1 ml of this solution with 999 ml of distilled water to get 200 ng per ml and finally, dilute 1 ml of this solution with 999 ml of Folic Acid Buffer A to get a standard solution containing 0.2 ng folic acid per ml. use 0.0, 0.5, 1.0, 2.0, 3.0, 4.0 and 5.0 ml per assay tube. Extreme care should be taken to avoid contamination of media or glassware used for the assay. Detergent free clean glassware should be used. Even small amount of contamination by foreign material can be lead to erroneous results.

## **COMPOSITION**

Ingredients	Gms / Ltr	
Vitamin free Casein acid hydrolysate	10.000	
L-Asparagine	0.600	
L-Tryptophan	0.200	
L-Cystine hydrochloride	0.760	
Dextrose (Glucose)	40.000	
Adenine sulphate	0.010	
Guanine hydrochloride	0.010	
Uracil	0.010	











Xanthine	0.020		
p-Amino benzoic acid (PABA)	0.001		
Pyridoxine hydrochloride	0.004		
Thiamine hydrochloride	0.0004		
Calcium pantothenate	0.0008		
Nicotinic acid	0.0008		
Biotin	0.00002		
Riboflavin (Vitamin B2)	0.001		
Glutathione	0.0052		
Polysorbate 80	0.100		
Sodium citrate	52.000		
Dipotassium hydrogen phosphate	6.400		
Magnesium sulphate	0.400		
Manganese sulphate	0.020		
Sodium chloride	0.020		
Ferrous sulphate	0.020		

### **PRINCIPLE**

The medium consists of Vitamin Free Casein acid hydrolysate, Dextrose, L-cystine which provide the necessary organic carbon and nitrogen source. Magnesium and sulphates are a cofactor for many metabolic reactions. Sodium chloride in the medium maintains the osmotic balance. Folic Acid Assay Medium contains all the necessary nutrients for the growth of the test organism except folic acid. The medium contains nutrients like amino acids, carbohydrates, purine, pyrimidines, salts, and vitamins.

## **INSTRUCTION FOR USE**

- Dissolve 11.1 grams in 100 ml distilled water.
- Heat to boiling for 2-3 minutes. Agitate to dispense the slight precipitate evenly.
- For assay, dispense 5 ml medium into each assay tube (containing increasing amounts of standard or unknown) and make up the total volume to 10 ml per tube with distilled water.
- Sterilize by autoclaving at 15 psi pressure (121°C) for 5 minutes. Cool to 45-50°C. Satisfactory results are obtained with Folic acid at levels of 0, 2, 4, 6, 8, 10 nanograms per assay tube.

## **QUALITY CONTROL SPECIFICATIONS**

Appearance of Powder : Cream to yellow homogeneous free flowing powder.

Appearance of prepared medium : Light amber coloured, clear solution, which may have slight precipitate.

pH (at 25°C)  $: 6.7 \pm 0.2$ 

## **INTERPRETATION**

Cultural characteristics observed after incubation.













Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Incubation Temperature	Incubation Period
Enterococcus hirae	8043	50-100	Good Gradual increase in growth with increasing standard Folic acid concentration 0,2,4,6,8,10 ng per assay tube is recorded as equivalent increase in absorbance at 620nm	35-37°C	18-24 Hours

#### **PACKAGING:**

In pack size of 100 gm bottles.

#### **STORAGE**

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 10-25°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

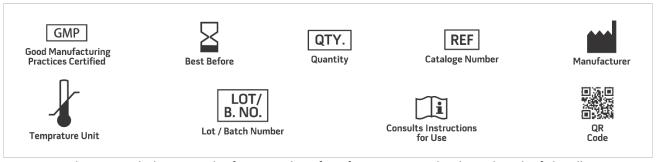
Product Deterioration: Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

#### **DISPOSAL**

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

## **REFERENCES**

- 1. Capps, Hobbs and Fox, 1948, J. Bacteriol., 55:869.
- 2. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
- 3. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th
- 4. Official Methods of Analysis of AOAC International, 2005, 19th Ed., Vol. II, Association of Analytical Chemists, Washington, D.C.



NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices. \*For Lab Use Only

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