

TM 1990 – BRAIN HEART INFUSION AGAR, MODIFIED (BHI AGAR MODIFIED)

INTENDED USE

For the cultivation of a wide variety of organisms like bacteria, yeasts and moulds.

PRODUCT SUMMARY AND EXPLANATION

Brain Heart Infusion Agar, Modified is similar to Brain Heart Infusion Agar with the exception of some of the medium constituents. Brain Heart Infusion Agar, Modified is highly nutritious and can support luxuriant growth of wide variety of microorganisms. It can be further enriched by the addition of blood or rendered selective by adding different antibiotics. It is a general purpose media used for primary isolation of aerobic bacteria from clinical specimens. Addition of 50 mg/l chloramphenicol or 40mg/l streptomycin or a mixture of 50mg/l gentamicin and 50mg/l chloramphenicol along with 5-10% sterile defibrinated blood is often recommended for inhibition of bacteria and isolation of pathogenic systemic fungi. A mixture of cycloheximide (0.5 g/l) and chloramphenicol (0.05 g/l) is also used for selective isolation of pathogenic fungi (incubation at 25-30°C for 1-2 weeks). Some fungi may be inhibited on this medium with 10% sheep blood, gentamicin and chloramphenicol.

COMPOSITION

Ingredients	Gms / Ltr		
Brain heart, infusion from (solids)	3.500		
Peptic digest of animal tissue	15.000		
Pancreatic digest of casein	10.000		
Dextrose	2.000		
Sodium chloride	5.000		
Disodium phosphate	2.500		
Agar	15.000		

PRINCIPLE

Peptones and infusions used in the media serves as sources of carbon, nitrogen, vitamins, amino acids, along with essential growth factors. Dextrose is the energy source. Sodium chloride maintains the osmotic equilibrium of the medium while disodium phosphate buffers the medium. Defibrinated sheep blood added to the basal medium provides essential growth factors for the more fastidious fungal organisms.

INSTRUCTION FOR USE

- Dissolve 53 grams in 1000 ml distilled water.
- Heat to boiling to dissolve the medium completely.
- Sterilize by autoclaving at 15 psi pressure (121°C) for 15 minutes.
- It may be further enriched with addition of blood.
- For addition cool the medium at 45-50°C and aseptically add 5% v/v sterile defibrinated sheep blood.
- Mix well and pour into sterile Petri plates.

QUALITY CONTROL SPECIFICATIONS















Appearance of Powder : Light yellow to yellow homogeneous free flowing powder.

Appearance of prepared medium : Basal medium: Light amber coloured, clear to slightly opalescent gel. After

addition of 5% v/v sterile defibrinated blood: Cherry red coloured, opaque gel

forms in Petri plates.

pH (at 25°C) : 7.4±0.2

INTERPRETATION

Cultural characteristics observed after incubation. (If desired add 5%v/v sterile defibrinated blood).

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Growth w/ blood	Recovery w/ blood	Incubation Temperature	Incubation Period
Escherichia coli	25922	50-100	Luxuriant	>=70%	Luxuriant	>=70%	35-37°C	18-24 Hours
Streptococcus pyogenes	19615	50-100	Luxuriant	>=70%	Luxuriant	>=70%	35-37°C	18-24 Hours
Staphylococcus aureus	25923	50-100	Luxuriant	>=70%	Luxuriant	>=70%	35-37°C	18-24 Hours
Streptococcus pneumoniae	6303	50-100	Luxuriant	>=70%	Luxuriant	>=70%	35-37°C	18-24 Hours

PACKAGING:

In pack size of 500 gm bottles.

STORAGE

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

Product Deterioration: Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

DISPOSAL

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES

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- 4. Creitz and Puckett, 1954, Am. J. Clin. Pathol., 24:1318.
- 5. Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Yolken R. H., (Eds.), 2003, Manual of Clinical Microbiology, 8th Ed., American Society for Microbiology. Washington. D.C.
- 6. Ajello L., Georg L., Kaplan W. and Kaufman L., 1963, CDC Laboratory Manual for Medical Mycology, PHS Publication No. 994, U.S. Govt. Office, Washington, D.C.





































NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices. *For Lab Use Only Revision: 08 Nov., 2019







