

# TM 1934 – BURKHOLDERIA CEPACIA AGAR BASE

#### **INTENDED USE**

For isolation of Burkholderia cepacia from the respiratory secretions of patients with cystic fibrosis and other nonclinical specimens.

#### PRODUCT SUMMARY AND EXPLANATION

Burkholderia cepacia is an important opportunistic pathogen and causes pulmonary infection among individuals with cystic fibrosis (CF). The organism may lead to Burkholderia cepacia syndrome, a neutralizing pneumonia associated with fever that culminates in to a rapid and fatal clinical deterioration. B. cepacia is difficult to isolate on routinely used laboratory media like MacConkey Agar, since B. cepacia is a slow grower and therefore it is usually outgrown by the faster growing Escherichia coli, Staphylococcus aureus, and Pseudomonas aeruginosa . Burkholderia Cepacia Agar is based on PC medium, which was originally devised by Gilligan. This medium was found to be superior to MacConkey Agar for growth of B. cepacia. The medium is made selective for B. cepacia by the incorporation of bile salts, crystal violet and antibiotics. The antibiotics included are Polymyxin B, Gentamycin, Ticarcillin in the form of freeze dried supplement (FD). The antibiotics (FD) namely ticarcillin, polymixin B and gentamycin inhibit gramnegative bacteria. B. cepacia metabolizes pyruvate forming alkaline end products. These end products elevate the pH of the medium. The phenol red indicator changes colour from pink orange to pink red in alkaline pH.

### **COMPOSITION**

Ingredients	Gms / Ltr	
Peptone	5.000	
Yeast extract	4.000	
Sodium pyruvate	7.000	
Potassium dihydrogen phosphate	4.400	
Disodium hydrogen phosphate	1.400	
Bile salts	1.500	
Ammonium sulphate	1.000	
Magnesium sulphate	0.200	
Ammonium ferrous sulphate	0.010	
Phenol red	0.020	
Crystal violet	0.001	
Agar	12.000	

#### **PRINCIPLE**

Peptone and yeast extract in the medium provides the nitrogenous, vitamin B source and other essential nutrients. Crystal violet, bile salts and antimicrobial agents are used as selective agents. Crystal violet and bile salts inhibits grampositive cocci including Enterococci and Staphylococci.

### **INSTRUCTION FOR USE**

- Dissolve 36.52 grams in 1000 ml distilled water.
- Heat to boiling to dissolve the medium completely.
- Sterilize by autoclaving at 15 psi (121°C) for 15 minutes.











Cool to 45-50°C and aseptically add the rehydrated contents of 2 vial of Burkholderia Selective Supplement (TS 323)

Mix well and pour in sterile Petri plates.

#### **QUALITY CONTROL SPECIFICATIONS**

**Appearance of Powder** : Light yellow to pink homogeneous free flowing powder.

**Appearance of prepared medium** : Yellow coloured clear to slightly opalescent gel forms in Petri plates.

pH (at 25°C) : 6.2±0.2

### **INTERPRETATION**

Cultural characteristics observed after incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Colour of colony	Incubation Temperature	Incubation Period
Burkholderia cepacia	25608	50-100	Good- luxuriant	>=50%	Sage green colonies with bright pink medium	35-37°C	≤48 Hours
Burkholderia cepacia	25416	50-100	Good- luxuriant	>=50%	Sage green colonies with bright pink medium	35-37°C	≤48 Hours
Pseudomonas aeruginosa	9027	>=10³	Inhibited	0%	-	35-37°C	72 Hours

# PACKAGING:

In pack size of 500 gm bottles.

# **STORAGE**

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

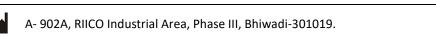
**Product Deterioration:** Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

## **DISPOSAL**

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

# **REFERENCES**

- 1. Whitby P. W., 1998, J. Clin. Microbiol., 36:1642 1645
- 2. Gilligar, Gage, Bradshaw, schidlow and Deciscoo, 1985, J. Clin. Microbiol., 22:5.
- 3. MacDonald Gilligan, Welch, Reller and Menegus, 1994, Vol. 5:1, Cystic Fibrosis Foundation, Washington, D.C.
- 4. Gilligan, 1996. Clin. Microbiol. Newsl. 18:83.
- 5. Christensen et al, 1980, J. Clin. Microbiol., 27:270.

































**NOTE:** Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices. \*For Lab Use Only

Revision: 28 Oct., 2023







