

THTS 012– TRANSPORT SWABS W/ CARY BLAIR MEDIUM

INTENDED USE

For recovery of aerobic, anaerobic and fastidious bacteria from faecal specimen.

PRODUCT SUMMARY AND EXPLANATION

Transport Medium is a non-nutritive, chemically defined, semisolid, buffered medium. The sole purpose of this medium is to maintain the viability of organisms during the time from collection to examination of the specimen. Transport Medium should be essentially non-nutritive so that the test organisms do not increase in numbers during transport. Transport media were originally formulated by Stuart et al for carrying gonococcal specimens to the laboratory, Cary and Blair devised a new medium containing fewer nutrients, low oxidation-reduction potential and a high pH. Cary-Blair Medium w/o Charcoal is used for collection and transport of clinical specimens. It is also recommended by APHA and various authors for transport of specimens. Since this transport media has a high pH, viability of *Vibrio* cultures can be maintained for a longer duration. This medium also facilitates the recovery of *Salmonella* and *Shigella* species.

COMPOSITION

Ingredients	Gms / Ltr
Agar	5.000
Sodium chloride	5.000
Sodium thioglycollate	1.500
Disodium phosphate	1.100

PRINCIPLE

Cary-Blair Medium Base is prepared with minimal nutrients to facilitate survival of organisms without multiplication. Sodium thioglycollate provides a low oxidation-reduction potential. Alkaline pH of the medium minimizes bacterial destruction due to the formation of acid. The sodium chloride and calcium chloride levels help control cell permeability and provide an osmotically balanced environment for the preservation of viable bacterial cells. Disodium hydrogen phosphate helps maintain a stable pH and prevents pH fluxes that may be detrimental to the organisms present in clinical specimens.

Note: The specimen should be inoculated in suitable medium as soon as possible and must not be kept at room temperature for more than 24 hours. Some contaminants may also grow, if specimen is kept for longer period in transport medium.

INSTRUCTION FOR USE

1. Use the medium, provided along with the swab to collect and transport the microbiological sample.
2. Collect the sample with the sterile swab and insert the capped swab with the sample till the bottom of the medium. Tighten the cap firmly
3. The sample and viability of organism(s) will be maintained during transportation.
4. After the transportation, the specimen should be inoculated in proper medium as soon as possible.

QUALITY CONTROL SPECIFICATIONS

Appearance	:	Colourless, clear to slightly opalescent gel
pH (at 25°C)	:	8.4 ±0.2
Sterility Check	:	Passes release criteria



INTERPRETATION

Culture characteristics observed after incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Recovery on SCDA	Incubation Temperature	Incubation Period
<i>Neisseria meningitidis</i>	13090	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Escherichia coli</i>	25922	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Vibrio cholerae</i>	15748	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Salmonella</i> Typhimurium	14028	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Enterobacter aerogenes</i>	13048	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Shigella flexneri</i>	12011	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Klebsiella pneumoniae</i>	13883	50-100	Good-Luxuriant	35-37 °C	18-72 Hours
<i>Vibrio parahaemolyticus</i>	15748	50-100	Good-Luxuriant	35-37 °C	18-72 Hours

PACKAGING:

In pack size of 50 No.

STORAGE

On receipt, store ready-to-use disposable swabs in the dark at 10 to 25° C. Avoid freezing and overheating. The medium may be used up to the expiration date and incubated for the recommended incubation times.

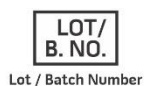
Product Deterioration: Do not use product if they show evidence of microbial contamination, discoloration, or any other signs of deterioration.

DISPOSAL

After use, prepared media, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES

1. Stuart, Toshach and Pastula, 1954 Can. J. Public Health, 45:73.
2. Cary and Blair, 1964, J. Bacteriol., 88:96.
3. Vanderzant C. and Splittstoesser D. F., (Eds.), 1992, Compendium of Methods for the Microbiological Examination of Foods, 3rd Ed., APHA, Washington, D.C.
4. Cary, Mathew, Fusillo and Harkins, 1965, Am. J. Clin. Pathol., 43:294
5. Gaines et al, 1965, Am. J. Trop. Med. Hyg. 14:136.
6. Morris and Heck, 1978.J. Clin. Microbiol.,8:616.
7. Leber, A.2016 Clinical Microbiology Procedures Handbook 4th edition 2016, ASM, Washington DC



NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.

*For Lab Use Only
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