

**BISMUTH SULPHITE AGAR**
**TM 039**

For selective isolation of Salmonellae from faeces, urine, sewage and other materials

**Composition**

Ingredients	Gms/Ltr.
Agar	20.00
Peptone	10.00
Bismuth sulphite	8.00
Beef extract	5.00
Dextrose	5.00
Disodium phosphate	4.00
Ferrous sulphate	0.30
Brilliant Green	0.025

\* Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers below 25°C and protect from direct Sunlight.

**Instructions for Use**

Dissolve 52.3gms in 1000ml of distilled water. Gently heat to boiling with gentle swirling and dissolve the medium completely. **DO NOT AUTOCLAVE**. Cool to 45-50°C prior to dispense.

**Appearance:** Greenish yellow colour, solution opalescent with flocculent precipitate  
**pH (at 25°C):** 7.7 ± 0.2

**Principle**

**BISMUTH SULPHITE AGAR** is used for the selective isolation of *Salmonella* spp. *Salmonella* constitute the most taxonomically complex group of bacteria among Enterobacteriaceae. Bismuth Sulphite Agar is a modification of **Wilson** and **Blair** formula. Also this medium favors use of larger inoculum as compared to other selective media, as it has unique inhibitory action towards gram-positive organisms and coliforms. Peptone and Beef extract serve as sources as carbon, nitrogen, vitamins and essential growth factors. Disodium phosphate acts as a buffering agent. Dextrose is a fermentable carbohydrate source of the growth of microorganisms. Bismuth Sulphite Indicator and Brilliant Green are complementary, inhibiting to Gram-positive bacteria and coliforms, allowing *Salmonella* spp. to grow. Ferrous Sulphate is used for H<sub>2</sub>S production. When H<sub>2</sub>S is present, the iron incorporated in medium is precipitated, and positive cultures produce the characteristic brown to black color with metallic sheen. Agar is the solidifying agent.

**Interpretation**

Cultural characteristics observed after inoculating (10<sup>3</sup> - 10<sup>5</sup>CFU/ml), on incubation at 35-37°C for 40 - 48 hours.

Microorganisms	ATCC	Inoculum (CFU/ml)	Growth	Recovery (%)	Appearance of colony
<i>Salmonella enteritidis</i>	13076	10 <sup>3</sup> - 10 <sup>5</sup>	Good-luxuriant	>=50%	Black with metallic sheen
<i>Salmonella typhi</i>	6539	10 <sup>3</sup> - 10 <sup>5</sup>	Good-luxuriant	>=50%	Black with metallic sheen

## PRODUCT DATA SHEET

<i>Escherichia coli</i>	25922	10 <sup>3</sup>	None - poor	<=10%	Brown – green (depends on the inoculum density)
<i>Shigella flexneri</i>	12022	10 <sup>3</sup>	None - poor	<=10%	Brown
<i>Enterobacter aerogenes</i>	13048	10 <sup>3</sup>	None- Poor	<=10%	Brown – green

### References

1. Andrews, W. H., G. A. June, P. S. Sherrod, T. S. Hammack, and R. M Amaguana. *Salmonella*, p. 5.01-5.20. In FDA Bacteriological analytical manual, 8<sup>th</sup> ed. AOAC International, Gaithersburg, MD. (1995).
2. Wilson, W. J., and E. M. Blair. A combination of bismuth and sodium sulphite affording an enrichment and selective medium for the typhoid-paratyphoid groups of bacteria. J. Pathol. Bacteriol. 29:310. (1926).
3. Wilson, W. J., and E. M. Blair. Use of a glucose bismuth sulphite iron medium for the isolation of *B. typhosus* and *B. proteus*. J. Hyg. 26:374-391. (1927).
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5. Isenberg, H. D. (ed.). Clinical microbiology procedures handbook, vol. 1. American Society for Microbiology, Washington, D.C. (1992).
6. Vanderzant, C., and D.F. Splittstoesser (eds.). Compendium of methods for the microbiological examination of foods, 3<sup>rd</sup> ed. American Public Health Association, Washington, D.C. (1992).
7. United States Pharmacopeial Convention. The United States pharmacopeia, 23<sup>rd</sup> ed. The United States Pharmacopeial Convention, Rockville, MD. (1995).